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|--------------------------------------------------------------------|--------------------|---------------------------|
| BIOCYTE EDUCATION AND RESEARCH FOUNDATION ,<br>SANGLI, MAHARASHTRA |                    |                           |
| Standard operating procedures of cell line                         |                    | Revision No: 01           |
|                                                                    |                    | Date of Issue: 21/07/2024 |
| Cell biology Lab.                                                  | SOP NO- BiRD/CBL/1 | Effective Date:21/07/2024 |

| Sr no.                  | Cell line  | <u>Cell line Name</u>                                    |
|-------------------------|------------|----------------------------------------------------------|
| <b>Cancer Cell Line</b> |            |                                                          |
| 1                       | MCF-7      | Human; Mammary gland; Breast; Adenocarcinoma cell line   |
| 2                       | MDA-MB-231 | Human; Adenocarcinoma; Mammary gland cell line           |
| 3                       | HepG2      | Human; Liver Cancer cell line                            |
| 4                       | COLO 320DM | Human Colon Cancer cell line                             |
| 5                       | COLO 205   | Human Colon Cancer cell line                             |
| 6                       | PC-3       | Human; Adenocarcinoma; Prostate cell line                |
| 7                       | A549       | Human; Lung cancer cell line                             |
| 8                       | B16-F10    | Mouse; Skin; Melanoma cell line                          |
| 9                       | HL-60      | <u>Human Leukemia</u> cell line                          |
| 10                      | MG-63      | Human; Osteosarcoma; Bone cell line                      |
| 11                      | SH-SY-5Y   | Human: Brain; Neuroblastoma cell line                    |
| 12                      | SKOV3      | Human; Ovary Adenocarcinoma cell line                    |
| 13                      | HCT 116    | Human Colorectal adenocarcinoma Cell line                |
| 14                      | HT-29      | Human Colorectal adenocarcinoma Cell line                |
| 15                      | CaCo2      | Immortalized Human colorectal Cell Line                  |
| <b>Normal Cell line</b> |            |                                                          |
| A                       | L929       | Connective Tissue; mouse, Mus Muscular                   |
| B                       | Min6       | Mouse Mus Musculus; Pancreatic B cells                   |
| C                       | RAW 264.7  | Murine Monocyte/ Macrophage- Anti-inflammatory cell line |

### **1) MCF-7**

#### **Human; Mammary gland; Breast; Adenocarcinoma**

Growth medium: Minimum essential medium (Eagle) with 2 mM L-glutamine and Earle's BSS adjusted to contain 1.5 g/L Na bicarbonate, 0.1 mM non-essential amino acids and 1 mM Na pyruvate and supplemented with 0.01 mg/ml calf insulin, 90%; fetal calf serum, 10%

Growth conditions: Temperature- 37°C; Carbon-dioxide 5% atmosphere

Remove medium, and rinse with TPVG solution. Remove the solution and add an additional 1 to 2 ml of TPVG solution. Allow the flask to sit at room temperature until cells detach. Add fresh culture medium, aspirate and dispense into new culture flasks.

Split ratio of 1:3 is suggested

Medium change: 3 times per week

### **2) MDA-MB-231**

#### **Human; Adenocarcinoma; Mammary Gland**

Growth Medium: Leibovitz's L-15 medium with 2 mM L-glutamine, 90%; fetal bovine serum 10%

Growth Condition: Temperature 37°C; No Carbon-dioxide 5% atmosphere

Remove growth medium from flask, Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense into new flask.

Recommended Split ratio: 1:2

Medium Change: 3 times per week

### **3) HepG2**

#### **Human; Liver cancer cell line**

Growth Medium: Minimum essential medium (Eagle) with 2 mM L- glutamine and Earle's BSS adjusted to contain 1.5 g/L Na bicarbonate, 0.1 mM non-essential amino acids and 1.0 mM Na pyruvate, 90% fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:4 is recommended

Medium Change: Twice per week

#### **4) COLO 320DM**

##### **Human; Colon cancer cell line**

Growth Medium: RPMI 1640 medium adjusted to contain 1.5 g/L Na bicarbonate, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:5 recommended

Medium Change: 1 to 2 times per week

#### **5) COLO 205**

##### **Human; Colon cancer cell line**

Growth Medium: RPMI 1640 medium with 2 mM L-glutamine adjusted to contain 1.5 g/L Na bicarbonate, 4.5 g/L glucose, 10 mM HEPES and 1.0 mM Na pyruvate, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks

Split ratio of 1:3 is recommended

Medium Change: Every 2 to 3 days

## **6) PC-3**

### **Human; Adenocarcinoma; Prostate**

Growth medium: Ham's F12K medium with 2 mM L- glutamine adjusted to contain 1.5 g/L Na bicarbonate, 90%; fetal calf serum, 10%

Growth conditions: Temperature- 37°C; 5% Carbon-dioxide atmosphere

Remove medium, and rinse with TPVG solution. Remove the solution and add an additional 1 to 2 ml of TPVG solution. Allow the flask to sit at room temperature until cells detach. Add fresh culture medium, aspirate and dispense into new culture flasks

Split ratio of 1:3 to 1:6 is suggested

Medium change: 2 times per week

## **7) A549**

### **Human; Lung cancer cell line**

Dulbecco's modified Eagle's medium with 4 mM L-glutamine adjusted to contain 1.5 g/L Na bicarbonate and 4.5 g/L glucose, 90%; fetal calf serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove medium and rinse with fresh medium. Remove the PBS completely and add fresh TPVG solution. Allow the flask to sit at room temperature until the cells detach. Centrifuge the cell suspension at 1000 rpm for 10 minutes, resuspend the pellet in fresh medium, aspirate and dispense into new flasks

Split ratio of 1:3 is suggested

Medium Change: Every 2 to 3 days

## **8) B16-F10 Mouse; Skin; Melanoma**

Growth Medium: Dulbecco's modified Eagle's medium with 4 mM L- glutamine adjusted to contain 1.5 g/L Na bicarbonate and 4.5 g/L glucose, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:6 is suggested

Medium Change: 3 times per week

## **9) HL-60**

### **Human Leukemia**

Growth Medium: RPMI 1640 medium with 2 mM L-glutamine adjusted to contain 1.5 g/L Na bicarbonate, 4.5 g/L glucose, and 1.0 mM Na pyruvate, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Cultures can be maintained by the addition of fresh medium or partial replacement of spent medium with fresh medium. Initiate cultures at  $1 \times 10^5$  cells/ml and maintain up to cell density of  $1 \times 10^6$  cells/ml

Medium Change: Every 2 to 3 days

## **10) MG-63**

### **Human; Osteosarcoma; Bone**

Growth Medium: Minimum essential medium (Eagle) with 2 mM L- glutamine and Earle's BSS adjusted to contain 1.5 g/L Na bicarbonate, 0.1 mM non-essential amino acids and 1.0 mM Na pyruvate 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:4 is suggested

Medium Change: 2 times per week

## **11) SH-SY-5Y**

### **Human: Brain; Neuroblastoma**

Growth Medium: Ham's F12K medium with 2mM L-glutamine adjusted to contain 1.5 g/L Na bicarbonate, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; 5% Carbon-dioxide atmosphere

Cells grow as a mixture of floating and adherent cells. Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:3 is suggested

Medium Change: Every 3-4 days

## **12) SKOV3**

### **Human; Ovary Adenocarcinoma**

Growth Medium: McCoy's 5A 90%; fetal bovine serum 10%

Growth Condition: Temperature 37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask, Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense into new flask.

Recommended Split ratio: 1:3

Medium Change: Every 2 to 3 days

### **13) HCT-116**

#### **Human; Colon Adenocarcinoma**

Growth Medium: McCoy's 5a medium 90%; fetal bovine serum 10%

Growth Condition: Temperature 37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cell come off substratum. Aspirate with fresh medium and dispense into new flask.

Recommended Split ratio: 1:3 to 1:8

Medium Change: Every 2 to 3 days

### **14) HT-29**

Human; Colorectal adenocarcinoma

Growth Medium: Dulbecco's modified Eagle's medium with 4.5 g/L glucose adjusted to contain 1.5 g/L Na bicarbonate, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature 37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Split ratio of 1:3 to 1:8 is suggested

Medium Change: 3 times per week

### **15) CaCo-2**

Human; Adenocarcinoma

Growth Medium: Minimum essential medium (Eagle) with 2 mM L-

glutamine and Earle's BSS adjusted to contain 1.5 g/L Na bicarbonate, 0.1 mM non-essential amino acids and 1.0 mM Na pyruvate, 80%; fetal bovine serum, 20%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense into new flask.

Split ratio of 1:2 to 1:3 is suggested

Medium Change: 1 to 2 times per week

### **A) Min6**

Mouse Mus Musculus; Pancreatic B cells

Growth Medium: Dulbecco's Modified Eagle's medium, 90%; fetal bovine serum 10%

Growth Condition: Temperature 37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cells come off substratum. Aspirate with fresh medium and dispense cell suspension into new flasks.

Recommended Split ratio: 1:4

Medium Change: Every 2 to 3 days

### **B) L929**

**Connective Tissue; mouse, Mus Muscular**

Growth Medium: Dulbecco's Modified Eagle's medium, 90%; fetal bovine serum 10%

Growth Condition: Temperature 37°C; Carbon-dioxide 5% atmosphere

Remove growth medium from flask. Rinse with TPVG twice and remove as much TPVG as possible leaving enough so that a thin film is formed over the cell sheet. Keep flask in horizontal position for some time. Tap flask against palm of hand and cell come off substratum. Aspirate with fresh medium and dispense into new flask.

Recommended Split ratio: 1:2 to 1:8

Medium Change: Every 3rd day

### **C) RAW 264.7**

**Mouse: Abelson murine Leukemia virus induced tumor**

Growth Medium: Dulbecco's modified Eagle's medium with 4 mM L- glutamine adjusted to contain 1.5 g/L Na bicarbonate and 4.5 g/L glucose, 90%; fetal bovine serum, 10%

Growth Conditions: Temperature-37°C; Carbon-dioxide 5% atmosphere

Subcultures are prepared by scraping. Remove old medium, add fresh dislodge cells and dispense into new flasks

Medium change: 3 times per week

### List of Bacteria

|                                 | Gram positive strains        | Purpose                           |
|---------------------------------|------------------------------|-----------------------------------|
| <i>Staphylococcus aureus</i>    | ATCC no 6538                 |                                   |
| <i>Bacillus subtilis</i>        | ATCC no 6633                 |                                   |
|                                 | <b>Gram Negative strains</b> |                                   |
| <i>E.coli</i>                   | ATCC no 8739                 |                                   |
| <i>Pseudomonas aeruginosa</i>   | ATCC no 15442                |                                   |
|                                 | <b>Fungal strains</b>        |                                   |
| <i>Candida albicans</i>         | ATCC no 14053                |                                   |
| <i>Aspergillus niger</i>        | ATCC no 11414                |                                   |
| <i>Aspergillus brasiliensis</i> | ATCC no16404                 |                                   |
|                                 |                              |                                   |
|                                 | <b>Other Strain</b>          |                                   |
| <i>Propionibacterium acne</i>   | ATCC NO 11827                | Antiacne activity                 |
| <i>Malassezia furfur</i>        | ATCC no.14521-               | Antidandruff activity             |
| <i>Mycobacterium smegmatis</i>  | ATCC no. 101-                | Putative Anti Tubercular activity |